

WEST**Freeform Search**

Database:	US Patents Full-Text Database	▲
	US Pre-Grant Publication Full-Text Database	
	JPO Abstracts Database	
	EPO Abstracts Database	
	Derwent World Patents Index	
	IBM Technical Disclosure Bulletins	▼
Term:	termitomyces	
		▼
Display:	100	Documents in Display Format: CIT
		Starting with Number 1
Generate:	<input type="radio"/> Hit List <input checked="" type="radio"/> Hit Count <input type="radio"/> Image	

Search

Clear

Help

Logout

Interrupt

Main Menu

Show 8 Numbers

Edit 8 Numbers

Preferences

Search History

Today's Date: 12/12/2001

<u>DB Name</u>	<u>Query</u>	<u>Hit Count</u>	<u>Set Name</u>
USPT,PGPB	termitomyces	2	<u>L2</u>
USPT,PGPB	termitomyces clypeatus	0	<u>L1</u>

=> file biosis
COST IN U.S. DOLLARS

SINCE FILE	TOTAL
ENTRY	SESSION
0.42	0.42

FULL ESTIMATED COST

FILE 'BIOSIS' ENTERED AT 12:46:42 ON 12 DEC 2001
COPYRIGHT (C) 2001 BIOSIS(R)

FILE COVERS 1969 TO DATE.
CAS REGISTRY NUMBERS AND CHEMICAL NAMES (CNS) PRESENT
FROM JANUARY 1969 TO DATE.

RECORDS LAST ADDED: 5 December 2001 (20011205/ED)

The BIOSIS file has been reloaded. Enter HELP RLOAD and HELP REINDEXING for details.

=> termitomyces clypeatus
132 TERMITOMYCES
159 CLYPEATUS
L1 35 TERMITOMYCES CLYPEATUS
(TERMITOMYCES(W)CLYPEATUS)

=> d bib ab 1-35

L1 ANSWER 1 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	2001:473019	BIOSIS
DN	PREV200100473019	
TI	Hetero-aggregation with sucrase affects the activity, stability and conformation of extra- and intra-cellular cellobiase in the filamer fungus <i>T. clypeatus</i> .	
AU	Mukherjee, Sumana; Basak, Soumen; Khowala, Suman (1)	
CS	(1) Department of Applied Biochemistry, Indian Institute of Chemical Biology, 4 Raja S C Mullick Road, Calcutta, 700032: sumankhowala@iicb.res.in India	
SO	Enzyme and Microbial Technology, (September 5, 2001) Vol. 29, No. 4 213-224. print. ISSN: 0141-0229.	
DT	Article	
LA	English	
SL	English	
AB	Cellobiase (C) from <i>T. clypeatus</i> was found to be aggregated with sucrase (S) in extra- and intra-cellular fractions, when co-aggregates of the enzyme with sucrase with different activity ratios (C/S) were obtained during purification. The co-aggregates were compared for their activity, stability, and kinetic parameters with a purified sucrase-free cell preparation. The specific activity and stability of both the extra- and intra-cellular enzyme decreased significantly in the absence of sucrose. The catalytic activity (V_{max}/K_m) of sucrase-free cellobiase were decreased by 4236 and 652 fold compared to the crude enzyme in culture filtrate and mycelial extracts respectively. The stability of the enzyme also decreased versus pH, temperature and in the presence of chaotropic agents such as SDS, Gdn.HCl and urea after disaggregation from sucrase. Optimum temperatures of the free intra- and extra-cellular cellobiase were	

to 47degreeC from 45degreeC after the removal of sucrase from the co-aggregates, whereas optimum pH of the free enzyme and co-aggregates remained the same. Intra-cellular cellobiase had very high affinity sucrase and it was difficult to separate them. Cellobiase preparations from extra- and intra-cellular fractions were analysed by circular dichroism and light scattering spectroscopy and it was concluded that co-aggregation with sucrase was responsible for a change in conformation of cellobiase in the aggregates. The conformation of intra-cellular preparations were also different from those in the extra-cellular fractions. Instant regain of cellobiase activity in intra- and extra-cellular preparations were obtained on the addition in vitro sucrase from the respective fractions to the incubation mixture. The experiments suggested that hetero-aggregation with sucrase regulate activity and stability of cellobiase in the fungus.

L1 ANSWER 2 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 2000:164088 BIOSIS
 DN PREV200000164088
 TI Some records of Termitomyces from Old world Rainforests.
 AU Turnbull, E. (1); Watling, R. (1)
 CS (1) Royal Botanic Garden, Edinburgh, EH3 5LR UK
 SO Kew Bulletin., (1999) Vol. 54, No. 3, pp. 731-738.
 ISSN: 0075-5974.
 DT Article
 LA English
 SL English
 AB Eleven species of Termitomyces are enumerated from the rainforests Old world, seven from Malaysia and seven from the Republic of Cameroon with three species common to both. Lepiota discipes Henn. is synonymous with Termitomyces letestui and a second record is made of Tricholoma termitomycoides Corner, which resembles Termitomyces heimii Nataraj

L1 ANSWER 3 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1998:98960 BIOSIS
 DN PREV199800098960
 TI **Termitomyces clypeatus** controls secretion of extracellular amyloglucosidase by regulating exocytosis of vacuolar enzyme.
 AU Sengupta, Trishna; Hazra, P. P.; Mukhopadhyay, A.; Sengupta, S. (1)
 CS (1) Dep. Applied Biochem., Indian Inst. Chem. Biol., 4 Raja S. C. M Rd., Calcutta 70032 India
 SO FEMS Microbiology Letters, (Jan. 1, 1998) Vol. 158, No. 1, pp. 101-107.
 ISSN: 0378-1097.
 DT Article
 LA English
 AB An extracellular amyloglucosidase (56 kDa) of **Termitomyces clypeatus** which was accumulated intracellularly in absence of Krebs cycle acid dextrin medium, remained mostly inside the plasma membrane. The enzyme localised in mycelial vacuoles as a sucrase-amyloglucosidase aggregate. The intracellular pool contained both the free sucrase and the aggregate while vacuoles contained only aggregate without any free enzyme. The aggregate, partially purified to PAGE homogeneity, contained amyloglucosidase and sucrase in the ratio of 1:1. A cellular regulation influenced by the presence of Krebs cycle acids, was indicated to be

present at the level of exocytosis of vacuolar enzyme.

L1 ANSWER 4 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1998:53127 BIOSIS

DN PREV199800053127

TI Purification and characterization of an extracellular beta-xylosidase
Termitomyces clypeatus.

AU Bhattacharyya, Saswati; Khowala, Suman; Kumar, A.; Sengupta, S. (1)

CS (1) Dep. Applied Biochem., Indian Inst. Chem. Biol., 4 Raja S. C. M Road, Calcutta 700 032 India

SO Biotechnology Progress, (Nov.-Dec., 1997) Vol. 13, No. 6, pp. 822-8
ISSN: 8756-7938.

DT Article

LA English

AB *Termitomyces clypeatus* liberated beta-xylosidase (EC 3.2.1.37) optimally in xylan medium but poorly in cellulose medium. The enzyme activity reached 5-6% of that of xylanase liberated in xylan medium culture filtrate enzyme, purified 5-fold by ammonium sulfate precipitation, BioGel P-200, and DEAE-Sephadex anion exchange chromatographies at pH 5.0, was homogeneous (190 kDa) in polyacrylamide gel electrophoresis (PAGE) and in high-performance gel permeation chromatography (HPG PLC) but contained high amounts of cellobiase and sucrase and gave multiple protein bands in SDS-PAGE (SDS = sodium dodecyl sulfate). The aggregate was subsequently beta-xylosidase fractions decreasing sucrase contents. The sucrase free beta-xylosidase resolved by DEAE-anion exchange chromatography at pH 6.0 into a number of fractions subsequently purified to 55.6-fold by hydrophobic interaction chromatography on a phenyl-sepharose column. The enzyme was a homodimeric 94 kDa protein, both in SDS-PAGE and HPG PLC. The physicochemical properties of the enzyme were similar to those of other fungal beta-xylosidases, and the enzyme had no unrelated glycosidase activity. The purified (94 kDa) and aggregated forms (190 kDa) of beta-xylosidase had the same pH optima (5.0), temperature optima (60°C), substrate specificities, and sensitivities toward end product inhibition by xylan or to the actions of SDS, urea, and guanidine hydrochloride. But aggregated enzyme was reasonably stable in the pH and temperature range where purified enzyme was completely inactive. The protein-protein aggregation appeared to confer additional stability to the beta-xylosidase toward extracellular denaturing conditions.

L1 ANSWER 5 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1997:461021 BIOSIS

DN PREV199799760224

TI Regulation of protein secretion by mycelial culture of the mushroom
Termitomyces clypeatus.

AU Hazra, P. P.; Sengupta, Trishna; Mukhopadhyay, A.; Ghosh, A. K.; Mukherjee, M.; Sengupta, S. (1)

CS (1) Dep. Applied Biochemistry, Indian Inst. Chemical Biol., 4 Raja Mullick Rd., Calcutta 700032 India

SO FEMS Microbiology Letters, (1997) Vol. 154, No. 2, pp. 239-243.
ISSN: 0378-1097.

DT Article

LA English

AB **Termitomyces clypeatus** secreted a 24-kDa xylanase constitutively in xylan medium, but required a gluconeogenic amino acid or Krebs cycle for the secretion of a 56-kDa amyloglucosidase in dextrin medium. Aspartate, glutamate, succinate and fumarate all increased secreted amyloglucosidase from 50% to 90% and enzyme production by 10-fold. Little effect on xylanase production. Glutamate or succinate stimulated *in vitro* release of intracellular amyloglucosidase from washed mycelia in the presence of cycloheximide. Amyloglucosidase accumulated in the absence of glutamate was a high-molecular-mass protein that did not migrate in SDS-PAGE. Cellular regulation by the fungus of the secretion of amyloglucosidase was indicated.

L1 ANSWER 6 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1997:318733 BIOSIS

DN PREV199799609221

TI Characterisation of a xylanolytic amyloglucosidase of **Termitomyces clypeatus**.

AU Ghosh, Anil K.; Naskar, Amal K.; Sengupta, Subhobrata (1)

CS (1) Applied Biochemistry Dep., Indian Inst. Chemical Biol., 4, Raja Mullick Road, Calcutta-700 032 India

SO Biochimica et Biophysica Acta, (1997) Vol. 1339, No. 2, pp. 289-296
ISSN: 0006-3002.

DT Article

LA English

AB A xylanolytic amyloglucosidase of **Termitomyces clypeatus** was characterised with respect to other amyloglucosidases. The enzyme contained high alpha-helix destabilising amino acids but no sulphur acid. It contained high threonine and serine, analogous to other starch hydrolysing enzymes. Both xylanase and amyloglucosidase activities were gradually lost with the progress of tryptophan oxidation by NEM. Total inactivation occurred after oxidation of 4-5 tryptophan residues. In the presence of substrates (either starch or xylan), complete inactivation of either activities was not noticed even after oxidation of 7.7 mol tryptophan residues. Inactivation by HNBB was not possible in the absence of any denaturant. Only 4.9 mol of tryptophan could be modified in the presence of 5 M urea which resulted in only 42% inhibition of activity. Thus modified enzyme had higher V_m/K_m and lower pH optima in comparison to those of native enzyme. It was suggested that tryptophan was present at the substrate binding site and not at the active site. No such change in activity was noticed after modification of tyrosine, lysine or arginine residues. HPGPLC analysis of both dilute and concentrated enzyme samples indicated that the enzyme existed as an equilibrium mixture of monomer-oligomer. Perhaps for this reason molar mass of NAI modified enzyme appeared to be almost half of that modified by NAI in presence of substrate. Arrhenius plot of the enzyme also indicated reversible oligomerisation as a function of temperature.

L1 ANSWER 7 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1997:202849 BIOSIS

DN PREV199799502052

TI Acetyl esterase production by **Termitomyces clypeatus**.

AU Mukhopadhyay, A.; Harza, P. P.; Sengupta, T.; Ghosh, A. K.; Sengupta (1)

CS (1) Indian Inst. Chem. Biol., 4 Raja S.C. Mullick Road, Calcutta 70
India
SO Biotechnology Letters, (1997) Vol. 19, No. 2, pp. 159-161.
ISSN: 0141-5492.
DT Article
LA English
AB Production of acetyl esterase by *Termitomyces clypeatus* was stimulated by xylan, cellulose, arabinose and arabinose-containing polysaccharides in the growth medium. The culture filtrate was equally active with p-nitrophenyl acetate and acetyl xylan. Acetyl xylan was completely deacetylated by the enzyme. Activity was optimum at pH 6.5 and at 5 degree C. The Km values for p-nitrophenyl acetate and acetyl xylan 0.83 mM and 0.38% (w/v) with Vm of 48 and 55 mmole acetate produced cndot mg protein, respectively.

L1 ANSWER 8 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1996:274517	BIOSIS
DN	PREV199698830646	
TI	Regulation of endo-1,4-beta-glucanase secretion from <i>Termitomyces clypeatus</i> by carbon catabolic product(s).	
AU	Mukherjee, M.; Khowala, S.; Ghosh, A. K.; Sengupta, S. (1)	
CS	(1) Dep. Applied Biochem., Indian Inst. Chemical Biol., Calcutta-70 India	
SO	Folia Microbiologica, (1995) Vol. 40, No. 5, pp. 475-480. ISSN: 0015-5632.	
DT	Article	
LA	English	
AB	Secretion of CMCase by <i>Termitomyces clypeatus</i> was only observed in presence of a gluconeogenic amino acid, a citrate-cycle acid, maleate. Subinhibitory concentrations of glucosamine, or fluoride in the medium. The enzyme was not secreted in the presence of caffeine or IBMX or theophylline, and these phosphodiesterase inhibitors lowered the secretion of CMCase by glutamate. The presence of both glucosamine and glutamate in a cellulose medium were, however, antagonistic to CMCase secretion. In the growth medium, xylose and glucose were equivalent carbon source for the fungus while succinate was a poor source and strongly repressed growth at higher concentrations. Growth of <i>T. clypeatus</i> was highly favored in media containing xylose/glucose with succinate/glutamate. During growth of <i>T. clypeatus</i> in a glucose medium, the intracellular glucose level was stabilized by the presence of succinate, glutamate or glucosamine in the medium. All these observations suggested that a negative cellular feedback regulation, mediated by carbon catabolic product(s), existed in <i>T. clypeatus</i> which regulated the secretion of CMCase. A transient but significant increase of intracellular cAMP and cGMP levels was observed at the onset of mycelial growth in glucose and glucose/maleate media, respectively.	

L1 ANSWER 9 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1995:539832	BIOSIS
DN	PREV199598554132	
TI	Development of high-molar-mass cellobiase complex by spontaneous protein-protein interaction in the culture filtrate of <i>Termitomyces clypeatus</i> .	

AU Roy, S. B.; Ghosh, A. K.; Sengupta, S.; Sengupta, S. (1)
 CS (1) Dep. Applied Biochem., Indian Inst. Chemical Biol., Calcutta-70
 India
 SO Folia Microbiologica, (1994) vol. 39, No. 6, pp. 463-470.
 ISSN: 0015-5632.
 DT Article
 LA English
 AB The 450 kDa cellobiase from *Termitomyces clypeatus* which migrates as a single band on IEF, PAGE and SDS-PAGE, was found to possess appreciable sucrose activity. The fungus produced sucrose and cellobiase constitutively in different media but with different activity ratios. Kinetics of secretion of the two enzymes was similar under in vivo and in vitro conditions. HPGPLC analysis of the culture filtrates indicated the presence of both sucrose and cellobiase in the same protein fraction of different molar mass, even in the 30-kDa protein fraction. No free sucrose or cellobiase could be detected in the culture filtrates. It was also observed that fractionation of cellobiase by (NH₄)-2SO₄ precipitation was different with different amounts of associated sucrose activity present in the culture filtrate. The (NH₄)-2SO₄-precipitated cell fraction also contained cellobiases in proteins of widely varied molecular mass ranges. However, none of the low-molar mass proteins other than the 450-kDa enzyme could be purified, as all low-molar-mass fractions spontaneously aggregated to the 450-kDa enzyme. Hydrophobic chromatography of the (NH₄)-2SO₄-precipitated fractions followed by HPGPLC of the eluted active fraction yielded both cellobiase-free sucrose and a sucrose-containing cellobiase fraction. The cellobiase fraction, homogeneous in PAGE, was also a high-molar-mass protein complex dissociating into a number of protein bands on SDS-PAGE. It was suggested that the 450-kDa cellobiase was not liberated by the fungus as a pre-formed enzyme complex but that the complex developed through interaction of cellobiase with sucrose under in vitro conditions and the possibility of the involvement of other proteins in the aggregation cannot be excluded.

L1 ANSWER 10 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1995:391286	BIOSIS
DN	PREV199598405586	
TI	Purification and characterization of an amyloglucosidase from <i>Termitomyces clypeatus</i> that liberates glucose from xylan.	
AU	Ghosh, A. K.; Naskar, A. K.; Jana, M. L.; Khawala, S.; Sengupta, S.	
CS	(1) Indian Inst. Chem. Biol., 4, Raja S.C. Mullick Rd., Calcutta 700001, India	
SO	Biotechnology Progress, (1995) Vol. 11, No. 4, pp. 452-456. ISSN: 8756-7938.	
DT	Article	
LA	English	
AB	An amyloglucosidase was purified to homogeneity from the culture filtrate of <i>Termitomyces clypeatus</i> , using the following steps: ammonium sulfate fractionation, DEAE-Sephadex chromatography, and HP-GPLC on an Ultrahydrogel TSK-G3000 SWG column. The enzyme was a glycoprotein with a minimum molecular weight of 56 000. It had appreciable activity on glycogen, amylopectin, moderate activity on maltose, and little activity on starch. The enzyme, unlike fungal amyloglucosidase (<i>Aspergillus niger</i>), could not liberate glucose from xylans. The enzyme had $K_m = 1.81$ mg/mL and $v_{max} = 82.1$ μ -mol/min/mg for starch hydrolysis and $K_m = 4.36$ mg/mL and $v_{max} = 57.7$ μ -mol/min/mg for the hydrolysis of larch wood xylan. Among the	

different inhibitors, NBS and CDTA were the most potent. Previously enzyme was shown (Ehowala, S.; et al. Appl. Microbiol. Technol. 199 287-292) to have synergistic activity on xylan hydrolysis similar to xylanolytic enzymes: (alpha-arabinofuranosidase or (alpha-glucuronidase). Since the amyloglucosidase was not active on cellulose, arabinogalactan of glucose directly from xylan by the enzyme was indicated.

L1 ANSWER 11 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1995:320760	BIOSIS
DN	PREV199598335060	
TI	Short communication: simultaneous production of alpha-arabinofuranosidase and xylanase by <i>Termitomyces clypeatus</i> .	
AU	Sinha, N.; Sengupta, S. (1)	
CS	(1) Dep. Applied Biochem., Indiana Inst. Chem. Biol., 4 Raja S.C. M Road, Calcutta 700 032 India	
SO	world Journal of Microbiology & Biotechnology, (1995) Vol. 11, No. 359-360. ISSN: 0959-3993.	
DT	Article	
LA	English	
AB	<i>Termitomyces clypeatus</i> produced xylanase and alpha-L-arabinofuranosidase simultaneously in various media. The arabinofuranosidase had pH and temperature optima of 5.5 and 50 degrees respectively, and was stable at 50 degree C for 30 min and at pH values from 2 to 5. The partially purified enzyme was distinct from xylanase present in the same medium.	

L1 ANSWER 12 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1995:120194	BIOSIS
DN	PREV199598134494	
TI	<i>Termitomyces</i> of southeast Asia.	
AU	Pegler, D. N. (1); Vanhaecke, M.	
CS	(1) Herbarium, Royal Botanic Gardens, Kew, Richmond, Surrey TW9 3AE	
SO	Kew Bulletin, (1994) Vol. 49, No. 4, pp. 717-736. ISSN: 0075-5974.	
DT	Article	
LA	English	
AB	All species of the genus <i>Termitomyces</i> Heim (Agaricales, Pluteaceae) occur throughout southeast Asia and are collectively considered for the first time. Illustrated accounts for all the accepted species are provided together with a key to species.	

L1 ANSWER 13 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1993:303190	BIOSIS
DN	PREV199396021415	
TI	A hemolytic protein from cultured mycelia of mushroom, <i>Termitomyces clypeatus</i> .	
AU	Khowala, Suman; Banerjee, P. C.; Ghosh, A. K.; Sengupta, S. (1)	
CS	(1) Dep. Applied Biochem, Indian Inst. Chemical Biol., Calcutta 700 032 India	
SO	Indian Journal of Experimental Biology, (1993) Vol. 31, No. 1, pp.	

ISSN: 0019-5189.

DT Article

LA English

AB A hemolytic protein was purified from cultured mycelia of *Termitomyces clypeatus*. Some of the physico-chemical properties of the hemolysin studied. The protein was analyzed to be a lipoprotein and delipidated removed its hemolytic property. The monomeric protein subunit of the lipoprotein had a molecular weight of 64,000. Mode of action of the hemolysin were studied by observing protections of sugar and lipid components to hemolysin mediated lysis of red blood cells. It was concluded that the hemolysin possibly interacted with the phospholipid components of the blood cells causing lysis.

L1 ANSWER 14 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1993:124517 BIOSIS

DN PREV199395068617

TI Induced mutation of mycelial protoplast of mushroom, *Termitomyces clypeatus* for obtaining auxotrophic mutants.

AU Mukherjee, M.; Sengutpa, S. (1)

CS (1) Indian Inst. Chem. Biol., 4 Raja SC Mullick Rd., Calcutta 700 001 India

SO Indian Journal of Experimental Biology, (1992) vol. 30, No. 12, pp. 1206-1207.

ISSN: 0019-5189.

DT Article

LA English

AB *Termitomyces clypeatus*, which is aconidial as mycelial growth under laboratory condition was grown in submerged culture in the presence of N-methyl-N'-nitro-N-nitrosoguanidine (NTG). Protoplasts from the mycelia grown in the presence or in absence of NTG were irradiated for 10 min with UV light. Mycelia unexposed to NTG did not give any auxotrophic mutant. However, mutants characterized were found to be multiple auxotrophs.

L1 ANSWER 15 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1992:409290 BIOSIS

DN BA94:72490

TI SACCHARIFICATION OF XYLAN BY AN AMYLOGLucOSIDASE OF *TERMITOMYCES-Clypeatus* AND SYNERGISM IN THE PRESENCE OF XYLANASE.

AU KHOWALA S; GHOSH A K; SENGUPTA S

CS DEPT. APPLIED BIOCHEM., INDIAN INST. CHEMICAL BIOL., 4 RAJA S. C. MI ROAD, CALCUTTA 700032, INDIA.

SO APPL MICROBIOL BIOTECHNOL, (1992) 37 (3), 287-292.

CODEN: AMBIDG. ISSN: 0175-7598.

FS BA; OLD

LA English

AB An amyloglucosidase from a mycelial culture of the mushroom *Termitomyces clypeatus* hydrolysed larch wood xylan independently and synergistically with an endo- β (1 \rightarrow 4) xylanase of the same fungus. The glucoamylase saccharified xylan predigested with xylanase at a faster rate compared to that of xylanase acting on amylase-digested xylan. However, overall saccharification of xylan in both cases was the same. Only xylose was liberated from xylan by amylase digestion whereas xylose, xylitol and other oligosaccharides were liberated during xylanase digestion.

synergistic response of enzyme combinations was reflected in the liberation of glucose from xylan, rather than xylose. Glucoamylase xylanase activities on soluble and insoluble fractions of larch wood with different xylose and glucose contents suggested that synergism in xylanolysis by the presence of glucoamylase was dependent on the activity of the participating xylanase on the xylan preparation. It is suggested that possibly α -glucosidic linkages are present in xylan and that amyloglucosidase might be involved in xylanolysis.

L1 ANSWER 16 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1992:123771 BIOSIS
 DN BA93:69571
 TI SECRETION OF BETA GLUCOSIDASE BY TERMITOMYCES-CLYPEATUS REGULATION OF CARBON CATABOLITE PRODUCTS.
 AU KHOWALA S; SENGUPTA S
 CS DEP. APPLIED BIOCHEM., INDIAN INST. CHEMICAL BIOL., CALCUTTA 700 032 INDIA.
 SO ENZYME MICROB TECHNOL, (1992) 14 (2), 144-149.
 CODEN: EMTED2. ISSN: 0141-0229.
 FS BA; OLD
 LA English
 AB Casamino acids, irrespective of carbon source used, highly stimulate extracellular production of β -glucosidase by *T. clypeatus*, which produced very low amounts of enzyme in the presence of 1% (w/v) sucrose including xylose in minimal growth medium. Casamino acids did not effect the rate of sugar uptake or improve glucose transport by mushroom, they significantly reduced the intracellular/extracellular enzyme ratio by increasing the secretion of the enzyme from the cell pool into the filtrate. No phosphoenol pyruvate-mediated transport of sugars was detectable in the fungi. Few amino acids utilized as carbon source in mushroom and krebs cycle acids (poorly supporting growth) showed activities on enzyme production similar to that of casamino acids. Nonmetabolizable glucose analogue glucosamine also increased extracellular enzyme production. Liberation of enzyme from washed mycelia was stimulated by the presence of glutamate in the incubation mixture and was also to be insensitive to cycloheximide (30 μ g ml⁻¹). Regulation of the excretion of β -glucosidase in *T. clypeatus* by glucose catabolic product(s) was indicated.

L1 ANSWER 17 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1991:305269 BIOSIS
 DN BR41:13859
 TI REGULATION OF TERMITOMYCES-CLYPEATUS PROTOPLAST REGENERATION BY KREBS CYCLE ACIDS.
 AU MUKHERJEE M; SENGUPTA S
 CS INDIAN INST. CHEM. BIOL., 4 RAJA SC MULLICK ROAD, CALCUTTA 32, INDIA
 SO FEMS Microbiol. Lett., (1991) 80 (1), 41-44.
 CODEN: FMLED7. ISSN: 0378-1097.
 FS BR; OLD
 LA English

L1 ANSWER 18 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1991:292295 BIOSIS
 DN BA92:13310
 TI CULTURAL STUDIES ON THE GENUS TERMITOMYCES IN SOUTH AFRICA I. MACRO AND MICROSCOPIC CHARACTERS OF BASIDIOME CONTEXT CULTURES.
 AU BOTHA W J; EICKER A
 CS DEP. BOTANY, UNIV. PRETORIA, S. AFR. 0002.
 SO MYCOL RES, (1991) 95 (4), 435-443.
 CODEN: MYCRER. ISSN: 0953-7562.
 FS BA; OLD
 LA English
 AB Characteristics of basidiome context cultures of five South African species of Termitomyces were investigated, namely T. umkowaani, T. reticulatus, T. sagittaeformis, T. clypeatus and T. microcarpus. Microscopic characters of the cultures were very similar but macroscopic characters differed markedly. It was possible to distinguish between different species by relying strictly on macroscopic character. Growth characters did not change when the nutrient medium and incubation conditions were standardised, and proved to be a reliable taxonomic criterion for the species under investigation. With the exception of T. microcarpus, all the species produced conidiophores and holoarthritis conidia in culture with numerous aged, inflated, ungerminated conic (sphaerocysts). With the exception of T. clypeatus, conidiophores were aggregated into spherical, farinaceous sporodochia which resembled sporodochia. Conidiophores of T. clypeatus were closely compacted to synnematus structures. Cultures of T. microcarpus exhibited typical basidiomycetous growth characters. However, they differed significantly from cultures of the other species which, unlike T. microcarpus, formed conidiophores and produced a raised, tough, cerebriform mycelium mat which could be considered a stroma. It is suggested that T. microcarpus should be transferred to the genus Podabrella Singer.

L1 ANSWER 19 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1991:185673 BIOSIS
 DN BA91:100422
 TI PURIFICATION AND CHARACTERIZATION OF A BETA GLUCOSIDASE CELLOBIASE FROM MUSHROOM TERMITOMYCES-CLYPEATUS.
 AU SENGUPTA S; GHOSH A K; SENGUPTA S
 CS INDIAN INST. CHEM. BIOL., 4 RAJA S C MULLICK RD., CALCUTTA 700032,
 SO BIOCHIM BIOPHYS ACTA, (1991) 1076 (2), 215-220.
 CODEN: BBACQ. ISSN: 0006-3002.
 FS BA; OLD
 LA English
 AB A β -glucosidase with cellobiase activity was purified to homogeneity from the culture filtrate of the mushroom Termitomyces clypeatus. The enzyme had optimum activity at pH 5.0 and temperature 65°C and was stable up to 60°C and within pH 2-10. Among the substrates tested, p-nitrophenyl- β -D-glucopyranoside and cellobiose were hydrolysed best by the enzyme. K_m and V_m values for these substrates were 0.5, 1.25, 95, 91 $\mu\text{mol/min per mg}$, respectively. The enzyme had low activity towards gentiobiose, salicin and β -methyl-D-glucoside. Glucose and cellobiose inhibited the β -D-glucosidase (PNPGase) activity competitively with K_i of 1.7 and 1.9 mM, respectively. Molecular mass

the native enzyme was approximated to be 450 kDa by HPLC, whereas s dodecyl sulphate polyacrylamide gel electrophoresis indicated a mol mass of 110 kDa. The high molecular weight enzyme protein was prese intracellularly and extracellularly from the very early growth phas enzyme had a pI of 4.5 and appeared to be a glycoprotein.

L1 ANSWER 20 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN	1991:68789 BIOSIS
DN	BA91:37449
TI	ASSOCIATION OF TERMITOMYCES-SPP WITH FUNGUS GROWING TERMITES.
AU	GOWDA D K S; RAJAGOPAL D
CS	DEP. OF ENTOMOL., UNIV. OF AGRIC. SCI., GKVK, BANGALORE 560 065, IN
SO	PROC INDIAN ACAD SCI ANIM SCI, (1990) 99 (4), 311-316.
	CODEN: PIANDR. ISSN: 0253-4118.
FS	BA; OLD
LA	English
AB	Among 5 species of Termitomyces spp. associated with Odontotermes s Termitomyces microcarpus was the most dominant on the mound surface Odontotermes redemanni during the rainy season. This species was fc grow on the fungal comb fragments brought out by termites as the su for its growth. As a result, decrease in cellulose (5.9%), lignin (nitrogen (0.54%), carbon (11.2%), C:N ratio (1.37), crude fat (0.48 moisture (17.02%) and increase in ash content (20.15%) were observe was also observed that Termitomyces microcarpus was rich in proteir (39.16-43.37%) and mineral content.

L1 ANSWER 21 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN	1991:45914 BIOSIS
DN	BA91:24195
TI	SPECIES OF TERMITOMYCES OCCURRING IN SOUTH AFRICA.
AU	VAN DER WESTHUIZEN G C A; EICKER A
CS	DEP. BOT., UNIVERSITY PRETORIA, 0002 PRETORIA, SOUTH AFRICA.
SO	MYCOL RES, (1990) 94 (7), 923-937.
	CODEN: MYCRER. ISSN: 0953-7562.
FS	BA; OLD
LA	English
AB	Seven species of Termitomyces were identified-T. clypeatus, T. microcarpus, T. sagittiformis, T. schimperi, T. striatus, T. umkowa T. reticulatus sp. nov. The wood-destroying termite Odontotermes ba was found to be the most commonly associated termite species. Termi sagitiiformis was associated with Odontotermes latericius, a new re The morphologies of the Termitomyces spp. are described and illustr and their occurrence, distribution and termite associations are dis

L1 ANSWER 22 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN	1991:9222 BIOSIS
DN	BA91:9222
TI	REGULATION BY AMINO ACIDS OF ALPHA AMYLASE AND ENDO-BETA-1-4-GLUCAN INDUCTION IN MYCELIAL CULTURE OF THE MUSHROOM TERMITOMYCES-CLYPEATI
AU	SENGUPTA S; SENGUPTA S
CS	INDIAN INST. CHEM. BIOL., 4 RAJA S. C. MULLICK RD., CALCUTTA 700 03

INDIA.

SO CAN J MICROBIOL, (1990) 36 (9), 617-624.

CODEN: CJMIAZ. ISSN: 0008-4166.

FS BA; OLD

LA English

AB *Termitomyces clypeatus* constitutively liberated amyloglucosidase; liberation was not repressed by glucose. Growth of the mushroom in synthetic medium was slow with starch, and only amyloglucosidase was liberated. Yeast extract stimulated growth and enzyme production in medium, and α -amylase along with amyloglucosidase was detected extracellularly. The mushroom could not utilise cellulose or liberate endo- β (1 \rightarrow 4)-glucanase even when inducer cellobiose or glucose was added to cellulose at different concentrations. Cellobiose alone also failed to induce any extracellular endo- β (1 \rightarrow 4) glucanase production. Yeast extract in both cellulose and cellobiose supported liberation of endo- β (1 \rightarrow 4)-glucanase. Lactose was found to be a poor inducer even in yeast extract medium. However, α -amylase and endo- β (1 \rightarrow 4) glucanase were detected intracellularly at a basal level even when the enzymes were absent extracellularly under inducing and noninducing conditions. The intracellular enzymes were only freely liberated into the medium in presence of yeast extract. It appeared that induction of α -amylase and endo- β (1 \rightarrow 4)-glucanase was largely inhibited by the restricted liberation of the enzymes in absence of yeast extract. Catabolic inhibition observed at the late phase of enzyme production is proposed that catabolic inhibition might have a role in the enzyme liberation and that amino acids supported extracellular enzyme production by relieving this inhibition.

L1 ANSWER 23 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full
TextCiting
References

AN 1990:196921 BIOSIS

DN BA89:103592

TI BETA GLUCOSIDASE PRODUCTION BY THE MYCELIAL CULTURE OF THE MUSHROOM *TERMITOMYCES-CLYPEATUS*.

AU SENGUPTA S; SENGUPTA S

CS INDIAN INST. CHEM. BIOL., 4 RAJA S.C. MULLICK RD., CALCUTTA-700032,

SO ENZYME MICROB TECHNOL, (1990) 12 (4), 309-314.

CODEN: EMTED2. ISSN: 0141-0229.

FS BA; OLD

LA English

AB Extracellular cellobiase activity was detected in the mycelial culture of the mushroom *T. clypeatus* with different mono-, di-, and polysaccharides as carbon source. Higher carbohydrate (2-5%) in the medium strongly repressed enzyme production without inhibiting growth rates. On the other hand, nonglucose monosaccharides also could not improve extracellular enzyme activity. Casein hydrolysate (CH) in the medium at 1% (w/v) concentration largely improved enzyme titer irrespective of carbon source (glucose, xylose, cellobiose, starch) used. Extracellular activity appeared in high carbohydrate media in the presence of casein hydrolysate. The kinetics of extra- and intracellular production of the enzyme in cellobiose (CB) medium, with or without CH, indicated extracellular growth-dependent production of the enzyme. A maximum intracellular

of 8% of the total cellobiase was measured at the late phase of growth on CB medium. CH had no effect on pH, temperature optima, and thermal stability of the enzyme produced in different carbohydrate-containing media. *T. clypeatus* did not liberate any proteinase in the presence or absence of CH. Thus CH appeared not to improve enzyme titer by releasing any proteinase or stabilizing enzyme activity liberated in CH-free culture. It was therefore suggested that the constitutive production of cellobiase by *T. clypeatus* was under catabolic repression and CH probably released the repression to some extent. The β -glucosidase activity of the culture filtrate on p-nitrophenyl- β -D-glucose (pNPG), β -methyl-D-glucoside, and cellobiose had identical pH and temperature optima at 5° C and 65° C, respectively. The enzyme had higher affinity for aryl- β -D-glucose, while β -CH₃-D-glucoside was a very poor substrate for the enzyme. The activity of the enzyme was readily inhibited by glucose, whereas glucose analogues or any other related sugars did not have any appreciable inhibitory activity.

L1 ANSWER 24 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1989:181656	BIOSIS
DN	BA87:92922	
TI	ISOLATION AND REGENERATION OF PROTOPLASTS FROM <i>TERMITOMYCES-CLYPEATUS</i>	
AU	MUKHERJEE M; SENGUPTA S	
CS	INDIAN INST. CHEM. BIOL., 4 RAJA S. C. MULLICK ROAD, CALCUTTA 700 032, INDIA.	
SO	CAN J MICROBIOL, (1988) 34 (12), 1330-1332.	
	CODEN: CJMIAZ. ISSN: 0008-4166.	
FS	BA; OLD	
LA	English	
AB	A method for the efficient release of protoplasts from the mycelia of <i>Termitomyces clypeatus</i> and the conditions necessary for the regeneration of the protoplasts are described. It was possible to convert <i>T. clypeatus</i> protoplasts, to a concentration of 2×10^8 /mL of incubation mixture, by digesting the mycelia with a mixture of cell wall degrading enzymes, chitinase, and Novozym 234 for 3 h. Mycelial regeneration of the protoplasts was not detected in liquid regenerating medium, whereas more than 50% of the protoplasts developed into colonies on the same solid medium. Both direct hyphal growth and budding of the protoplasts with or without any hyphal development were observed on solid medium. However, budding of the protoplasts was only observed in the liquid regenerating medium.	

L1 ANSWER 25 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1988:460063	BIOSIS
DN	BA86:101782	
TI	CARBOXYMETHYLXYLAN A SPECIFIC SUBSTRATE DIRECTLY DIFFERENTIATING THE BACKBONE-HYDROLYZING AND SIDE CHAIN-REACTING BETA-D-1-4 XYLANASES OF THE MUSHROOM <i>TERMITOMYCES-CLYPEATUS</i> .	
AU	KHOWALA S; MUKHERJEE M; SENGUPTA S	
CS	DEPT. APPLIED BIOCHEM., INDIAN INST. CHEM. BIOL., 4 RAJA S C MULLICK ROAD, CALCUTTA 700032, INDIA.	
SO	ENZYME MICROB TECHNOL, (1988) 10 (9), 563-567.	
	CODEN: EMTE2. ISSN: 0141-0229.	
FS	BA; OLD	
LA	English	

AB The mushroom *Termitomyces clypeatus* produces two endoxylanases (D) (X) when grown in media containing dextrin and xylan as carbon source respectively. Endoxylanase (D) showed wide variation in its activity on different lots of xylan preparations, and its activity was found to be dependent upon the composition of xylyns. The xylose-liberating endoxylanase (X) did not discriminate between different xylyns. The activity of xylanase (D) was found to decrease as the proportion of xylose in different xylan preparations increased. The dialyzable oligosaccharide from the digestion of xylan by enzyme (D) contained constituent sugars xylan, whereas xylose was the main constituent sugar of the undialyzable fraction. Enzyme (D) also could not liberate any reducing group from carboxymethyl xylan (CMX), a suitable substrate for viscometric and colorimetric assays of endolytic activity of xylanases. CMX was found to be modified preferentially at the substituent sugars of xylan rather than at backbone residues. Thus CMX proved to be a specific substrate for colorimetric assay of true endoxylanase activity.

L1 ANSWER 26 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1987:384172	BIOSIS
DN	BA84:70669	
TI	MULTISUBSTRATE SPECIFIC AMYLASE FROM MUSHROOM <i>TERMITOMYCES-CLYPEATUS</i>	
AU	GHOSH A K; SENGUPTA S	
CS	INDIAN INST. CHEM. BIOL., 4 RAJA S. C. MULLICK ROAD, JADAVPUR, CALCUTTA 700 032, INDIA.	
SO	J BIOSCI (BANGALORE), (1987) 11 (1-4), 275-286. CODEN: JOBSDN. ISSN: 0250-4774.	
FS	BA; OLD	
LA	English	
AB	An amylase was purified from the culture filtrate of <i>Termitomyces clypeatus</i> by ammonium sulphate precipitation, DEAE-Sephadex chromatography and gel filtration on Bio-Gel P-200 column. The electrophoretically homogeneous preparation also exhibited hydrolytic activity (in a decreasing order) on amylose, xylan, amylopectin, galactarabinogalactan and arabinoxylan. The enzyme had characteristically endo-hydrolytic activity on all the substrates tested and no xylose, glucose, arabinose or glucuronic acid could be detected even after prolonged enzymatic digestion of the polysaccharides. Interestingly, the enzyme had similar pH optima (5.5), temperature optima (55° C), pH stability (pH 3-10) and thermal denaturation kinetics when acted on starch and xylan (larch wood). Km values were found to be 2.63 mg/nM for amylase and 6.25 mg/ml for xylanase activity. Hill's plot also indicated that the enzyme contained a single active site for both activities. EDTA was found to be most potent inhibitor. Ca ²⁺ , a common activator for amylase activity, appeared to be an inhibitor for this enzyme. Thus it appeared that the enzyme had multisubstrate specificity acting as α -amylase on starch and also acting as xylanase on side chain oligosaccharides of xylan containing α -linked sugars.	

L1 ANSWER 27 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1987:91994	BIOSIS
DN	BR32:41795	
TI	NOTES ON THE GENUS <i>TERMITOMYCES</i> HEIM IN MALAWI.	
AU	MORRIS B	

CS GOLDSMITHS COLL., UNIV. LONDON.
 SO SOC MALAWI J, (1986) 39 (1), 40-49.
 CODEN: SMJODY.
 FS BR; OLD
 LA English

L1 ANSWER 28 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1985:438076 BIOSIS
 DN BA80:108068
 TI AN INDUCIBLE XYLANASE OF THE MUSHROOM *TERMITOMYCES-CLYPEATUS* DIFFEI FROM THE XYLANASE-AMYLASE PRODUCED IN DEXTRIN MEDIUM.
 AU MUKHERJEE M; SENGUPTA S
 CS INDIAN INST. CHEM. BIOL., 4 RAJA S.C. MULLICK ROAD, JADAVPUR, CALCUTTA 700 032, INDIA.
 SO J GEN MICROBIOL, (1985) 131 (8), 1881-1886.
 CODEN: JGMIAN. ISSN: 0022-1287.
 FS BA; OLD
 LA English
 AB The mushroom *T. clypeatus* produces a single endoxylanase (1,4- β -D-xylan xylanohydrolase, EC 3.2.1.8) in the presence of either dextrin or xylan as sole source of C. The enzymes produced in the 2 conditions are different. The enzyme induced by xylan was purified from the culture filtrate of *T. clypeatus*. The enzyme preparation gave a single protein band on sodium dodecyl sulfate-polyacrylamide gel electrophoresis, corresponding to a MW of ~ 24,000. The enzyme has an isoelectric point at pH 4.0 and acts on arabinoxylan and arabinogalactan, but not amylopectin or galactomannan. It shows maximum activity on xylan (1,4- β -linked-D-xylopyranose units) at pH 3.5 and 55° C and is fairly stable up to 60° C. The Km for xylan is 4 mg ml⁻¹. Hg²⁺, Fe²⁺ and Ag⁺ are the most potent inhibitors of the enzyme. The pH optimum and MW of this inducible xylanase differ from those of the enzyme produced by the same organism grown in dextrin medium.

L1 ANSWER 29 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1984:78567 BIOSIS
 DN BR26:78567
 TI SACCHARIFICATION OF UNTREATED AGRO WASTES DURING MYCELIAL GROWTH OF MUSHROOM *TERMITOMYCES-CLYPEATUS* ON SOLID BEDS.
 AU SENGUPTA S; NASKAR A K; JANA M L
 CS DEP. APPLIED BIOCHEM., INDIAN INST. CHEM. BIOL., 4 RAJA S. C. MULLICK ROAD, CALCUTTA 700 032, INDIA.
 SO Biotechnol. Bioeng., (1984) 26 (2), 188-190.
 CODEN: BIBIAU. ISSN: 0006-3592.
 FS BR; OLD
 LA English

L1 ANSWER 30 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1983:214137 BIOSIS
 DN BA75:64137
 TI HEM AGGLUTINATING ACTIVITY IN EXTRACTS OF MYCELIA FROM SUBMERGED CULTURES.

AU BANERJEE P C; GHOSH A K; SENGUPTA S
 CS INDIAN INSTITUT CHEMICAL BIOL., CALCUTTA 700 032, INDIA.
 SO APPL ENVIRON MICROBIOL, (1982) 44 (4), 1009-1011.
 CODEN: AEMIDF. ISSN: 0099-2240.
 FS BA; OLD
 LA English
 AB Extracts from mycelia of 7 different mushrooms [Volvariella volvacea
Termitomyces clypeatus, *Panafolus papillionaceus*, *Gymnopilus*
chrysimyces, *Lentinus squarrosulus*, *Coprinus lagopus*, *C. altramenta*
 agglutinated erythrocytes of several species [sheep, guinea pig, ra
 mouse, goat, human]. More than 1 agglutinating factor was identifie
 the extracts of 3 different mycelia. Agglutination was partially ir
 nonspecifically by high concentrations of glucose, galactose, manno
 fucose and rhamnose.

L1 ANSWER 31 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1983:175698	BIOSIS
DN	BA75:25698	
TI	NUTRITIVE VALUE OF SOME NIGERIAN EDIBLE MUSHROOMS.	
AU	OGUNDANA S K; FAGADE O E	
CS	DEP. MICROBIOL., UNIV. IFE, ILE-IFE, NIGERIA.	
SO	FOOD CHEM, (1982) 8 (4), 263-268. CODEN: FOCHDJ. ISSN: 0308-8146.	
FS	BA; OLD	
LA	English	
AB	Samples of <i>Termitomyces robustus</i> , <i>T. clypeatus</i> and <i>Pleurotus tuber-</i> <i>were</i> analyzed for their nutrient and toxic substances. The <i>Termitom</i> <i>spp.</i> contained as much as 31% proteins and ~ 32% carbohydrates, of which at least 26% were reducing sugars. <i>P. tuber-regium</i> contained protein and 18.6% carbohydrates; of which only ~ 2.9% were reducing sugars. There was little difference in their crude fiber and ash co while the fat content of <i>T. robustus</i> was a little higher than those other samples. The ascorbic acid content of each of the <i>Termitomyce</i> (10 and 14.3 mg%) was much higher than that of <i>Pleurotus sp.</i> (3.3 n All mushroom samples were low in HCN and oxalate contents.	

L1 ANSWER 32 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1983:63068	BIOSIS
DN	BR24:63068	
TI	TERMITOMYCES-CLYPEATUS COLLECTED FROM IRIOMOTE ISLAND OKINAWA JAPAI	
AU	OTANI Y; SHIMIZU D	
CS	DEPARTMENT OF BOTANY, TSUKUBA BOTANICAL GARDEN NATIONAL SCIENCE MUS IBARAKI PREFECTURE.	
SO	Bull. Natl. Sci. Mus., Ser. B (Tokyo), (1981 (RECD 1982)) 7 (4), 13 CODEN: BMBBD6. ISSN: 0385-2431.	
FS	BR; OLD	
LA	English	

L1 ANSWER 33 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

	Full Text	Citing References
AN	1982:51289	BIOSIS
DN	BR22:51289	

TI MODE OF ACTION OF AN ENDO XYLANASE ISOLATED FROM THE MUSHROOM
TERMITOMYCES-CLYPEATUS.
AU GHOSH A K
CS INDIAN INST. EXP. MED., CALCUTTA 700 032.
SO ANNUAL MEETING AND 2ND CONGRESS OF THE FEDERATION OF ASIAN AND OCEA
BIOCHEMISTS, BANGALORE, INDIA, DEC. 14-18, 1980. INDIAN J BIOCHEM E
(1981) 18 (4), 110.
CODEN: IJBBBQ. ISSN: 0301-1208.
DT Conference
FS BR; OLD
LA English

L1 ANSWER 34 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
--------------	----------------------

AN 1980:233622 BIOSIS
DN BA70:26118
TI PURIFICATION AND PROPERTIES OF XYLAN HYDROLASE EC-3.2.1.8 FROM MUSH
TERMITOMYCES-CLYPEATUS.
AU GHOSH A K; BANERJEE P C; SENGUPTA S
CS INDIAN INST. EXP. MED., 4 RAJA S.C. MULLICK RD., CALCUTTA 700 032,
BENGAL, INDIA.
SO BIOCHIM BIOPHYS ACTA, (1980) 612 (1), 143-152.
CODEN: BBACAQ. ISSN: 0006-3002.
FS BA; OLD
LA English
AB The endoxylanase (1,4- β -D-xylan xylanohydrolase, EC 3.2.1.8) from t
culture filtrate of a mushroom, *T. clypeatus*, was purified 93-fold
ammonium sulfate precipitation, ion-exchange chromatography
(DEAE-Sephadex) and gel permeation chromatography (Bio-Gel P-200).
enzyme preparation gave a single protein band on disc gel electroph
at pH 9.5, and has a MW of about 90,000. It acts on amylopectin,
arabinoxylan and arabinogalactan. The enzyme shows maximum activity
xylan (1,4- β -linked D-xylopyranose units) at pH 5.5 and at 55°
C and is fairly stable between pH 3 and 10 and temperatures up to
60° C. The Km is 4 mg of xylan/ml. Hg2+ is the most potent
inhibitor, whereas Fe2+, Ag+, iodoacetate and phosphate moderately
the enzyme activity.

L1 ANSWER 35 OF 35 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
--------------	----------------------

AN 1976:217306 BIOSIS
DN BA62:47306
TI SOUTH INDIAN AGARICALES PART 1 TERMITOMYCES.
AU NATARAJAN K
SO KAVAKA, 3 1975 (RECD 1976), 63-66.
CODEN: KVAKAH.
FS BA; OLD
LA Unavailable
AB Four species of *Termitomyces*, i.e., *T. badius* Otieno, *T. clypeatus*
T. microcarpus (Berk. et Br.) Heim and a new species, *T. indicus*, a
described.

=>
=>

=>
=>
=>
=>
=>
=>

YOU HAVE RECEIVED 8 CONSECUTIVE ARROW PROMPTS (=>)

The arrow (=>) is the system prompt, where you enter a command. For an explanation of system commands, files, formats, etc., enter "HELP" and the name of the item you want explained at an arrow prompt (=>). Enter "HELP COMMANDS" for a list of commands that can be used in this file. Enter "HELP MESSAGES" for a list of online explanations that are available. The "?" can be used as a synonym for "HELP".

Help is also available at any prompt, and after any error message. Enter "HELP" or "?" at a prompt to see an explanation of the options. After an error message, enter "HELP" or "?" at the next prompt and you will receive a more detailed explanation of the error and how to correct it.

Automatic help is also available. When AUHELP is 'ON', you will automatically receive help following an error message. For more information on AUHELP, enter "HELP SET AUHELP" at an arrow prompt (=>).

Users who need additional assistance can contact the Help Desk at their nearest STN Service Center. Enter "HELP STN" for information on STN Service Centers. You may also choose to contact the database representative for the file you are searching, for more detailed help on database content and search strategy. For information on how to contact database representatives for the current file, enter "HELP DESK" at an arrow prompt (=>).

IF YOU REQUIRE FURTHER HELP, PLEASE CONTACT YOUR LOCAL HELP DESK

=>
=>
=>
=>
=>

* * * * * RECONNECTED TO STN INTERNATIONAL * * * * *
SESSION RESUMED IN FILE 'BIOSIS' AT 12:56:32 ON 12 DEC 2001
FILE 'BIOSIS' ENTERED AT 12:56:32 ON 12 DEC 2001
COPYRIGHT (C) 2001 BIOSIS(R)
COST IN U.S. DOLLARS

	SINCE FILE ENTRY	TOTAL SESSION
FULL ESTIMATED COST	75.05	75.47

=> d his

(FILE 'HOME' ENTERED AT 12:45:43 ON 12 DEC 2001)

L1 FILE 'BIOSIS' ENTERED AT 12:46:42 ON 12 DEC 2001
35 TERMITOMYCES CLYPEATUS

=> cellobiase

447 CELLOBIASE
26 CELLOBIASES
L2 454 CELLOBIASE

(CELLOBIASE OR CELLOBIASES)

=> l1 and l2

L3 5 L1 AND L2

=> d bib ab 1-5

L3 ANSWER 1 OF 5 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 2001:473019 BIOSIS

DN PREV200100473019

TI Hetero-aggregation with sucrase affects the activity, stability and conformation of extra- and intra-cellular cellobiase in the filamer fungus *T. clypeatus*.

AU Mukherjee, Sumana; Basak, Soumen; Khowala, Suman (1)

CS (1) Department of Applied Biochemistry, Indian Institute of Chemical Biology, 4 Raja S C Mullick Road, Calcutta, 700032: sumankhowala@iicb.res.in India

SO Enzyme and Microbial Technology, (September 5, 2001) Vol. 29, No. 4 213-224. print. ISSN: 0141-0229.

DT Article

LA English

SL English

AB Cellobiase (C) from *T. clypeatus* was found to be aggregated with sucrase (S) in extra- and intra-cellular fractions, when co-aggregates of the enzyme with sucrase with different activity ratios (C/S) were obtained during purification. The co-aggregates were compared for their activity, stability, and kinetic parameters with a purified sucrase-free cellobiase preparation. The specific activity and stability of both extra- and intra-cellular enzyme decreased significantly in the absence of sucrase. The catalytic activity (V_{max}/K_m) of sucrase-free cellobiase preparations were decreased by 4236 and 652 fold compared to the crude enzyme in culture filtrate and mycelial extracts respectively. The stability of the enzyme also decreased versus pH, temperature and in the presence of chaotropic agents such as SDS, Gdn.HCl and urea after disaggregation with sucrase. Optimum temperatures of the free intra- and extra-cellular cellobiase were shifted to 47°C from 45°C after the removal of sucrase from the co-aggregates, whereas optimum pH of the free enzyme co-aggregates remained the same. Intra-cellular cellobiase had very high affinity for sucrase and it was difficult to separate them. Cellobiase preparations from extra- and intra-cellular fractions were analysed by circular dichroism and light scattering spectroscopy and it was concluded that co-aggregation with sucrase was responsible for a change in conformation of cellobiase in the aggregates. The conformation of intra-cellular enzyme preparations were also different from those of extra-cellular fractions. Instant regain of cellobiase activity in the *in vitro* of free sucrase from the respective fractions to the incubation mixture. The experiments suggested that hetero-aggregation with sucrase regulates the activity and stability of cellobiase in the fungus.

L3 ANSWER 2 OF 5 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1998:53127 BIOSIS

DN PREV199800053127
 TI Purification and characterization of an extracellular beta-xylosidase
Termitomyces clypeatus.
 AU Bhattacharyya, Saswati; Khawala, Suman; Kumar, A.; Sengupta, S. (1)
 CS (1) Dep. Applied Biochem., Indian Inst. Chem. Biol., 4 Raja S. C. M
 Road, Calcutta 700 032 India
 SO Biotechnology Progress, (Nov.-Dec., 1997) Vol. 13, No. 6, pp. 822-8
 ISSN: 8756-7938.
 DT Article
 LA English
 AB **Termitomyces clypeatus** liberated beta-xylosidase (EC 3.2.1.37)
 optimally in xylan medium but poorly in cellulose medium. The enzyme
 activity reached 5-6% of that of xylanase liberated in xylan medium
 culture filtrate enzyme, purified 5-fold by ammonium sulfate
 precipitation, BioGel P-200, and DEAE-Sephadex anion exchange
 chromatographies at pH 5.0, was homogeneous (190 kDa) in polyacryla
 gel electrophoresis (PAGE) and in high-performance gel permeation l
 chromatography (HPG PLC) but contained high amounts of **cellobiase** ar
 sucrase and gave multiple protein bands in SDS-PAGE (SDS = sodium c
 sulfate). The aggregate was subsequently beta-xylosidase fractions
 decreasing sucrase contents. The sucrase free beta-xylosidase resol
 DEAE-anion exchange chromatography at pH 6.0 into a number of fract
 subsequently purified to 55.6-fold by hydrophobic interaction
 chromatography on a phenyl-sepharose column. The enzyme was a homog
 94 kDa protein, both in SDS-PAGE and HPG PLC. The physicochemical
 properties of the enzyme were similar to those of other fungal
 beta-xylosidases, and the enzyme had no unrelated glycosidase activ
 The purified (94 kDa) and aggregated forms (190 kDa) of beta-xylosi
 had the same pH optima (5.0), temperature optima (60degreeC), subst
 specificities, and sensitivities toward end product inhibition by x
 or to the actions of SDS, urea, and guanidine hydrochloride. But
 aggregated enzyme was reasonably stable in the pH and temperature r
 where purified enzyme was completely inactive. The protein-protein
 aggregation appeared to confer additional stability to the beta-xyl
 toward extracellular denaturing conditions.

L3 ANSWER 3 OF 5 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
--------------	----------------------

AN	1995:539832 BIOSIS
DN	PREV199598554132
TI	Development of high-molar-mass cellobiase complex by spontaneous protein-protein interaction in the culture filtrate of Termitomyces clypeatus .
AU	Roy, S. B.; Ghosh, A. K.; Sengupta, S.; Sengupta, S. (1)
CS	(1) Dep. Applied Biochem., Indian Inst. Chemical Biol., Calcutta-70 India
SO	Folia Microbiologica, (1994) Vol. 39, No. 6, pp. 463-470. ISSN: 0015-5632.
DT	Article
LA	English
AB	The 450 kDa cellobiase from Termitomyces clypeatus which migrates a single band on IEF, PAGE and SDS-PAGE, was found to possess appre sucrase activity. The fungus produced sucrase and cellobiase constitutively in different media but with different activity ratio kinetics of secretion of the two enzymes was similar under in vivo vitro conditions. HPG PLC analysis of the culture filtrates indicate

presence of both sucrase and cellobiase in the same protein fraction of different molar mass, even in the 30-kDa protein fraction. No free sucrase or cellobiase could be detected in the culture filtrates. It was also observed that fractionation of cellobiase by (NH₄)-2SO₄ precipitation was different with different amounts of associated sucrase activity present in the culture filtrate. The (NH₄)-2SO₄-precipitated cellobiase fraction also contained cellobiases in proteins of wide varied molar mass ranges. However, none of the low-molar mass proteins other than the 450-kDa enzyme could be purified, as all low-molar mass fractions spontaneously aggregated to the 450-kDa enzyme. Hydrophobic chromatography of the (NH₄)-2SO₄-precipitated fractions followed by HPGPLC of the eluted active fraction yielded both cellobiase-free sucrase and a very low sucrase-containing cellobiase fraction. The cellobiase fraction, homogeneous in PAGE, was also a high-molar-mass protein complex dissociating into a number of protein bands on SDS-PAGE. It was suggested that the 450-kDa cellobiase was not liberated by the fungus as a preformed enzyme complex but that the complex developed through interaction of cellobiase with sucrase under *in vitro* conditions and the possibility of the involvement of other proteins in the aggregation cannot be excluded.

L3 ANSWER 4 OF 5 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1991:185673 BIOSIS

DN BA91:100422

TI PURIFICATION AND CHARACTERIZATION OF A BETA GLUCOSIDASE CELLOBIASE FROM A MUSHROOM TERMITOMYCES-CLYPEATUS.

AU SENGUPTA S; GHOSH A K; SENGUPTA S

CS INDIAN INST. CHEM. BIOL., 4 RAJA S C MULLICK RD., CALCUTTA 700032,

SO BIOCHIM BIOPHYS ACTA, (1991) 1076 (2), 215-220.

CODEN: BBACAQ. ISSN: 0006-3002.

FS BA; OLD

LA English

AB A β -glucosidase with cellobiase activity was purified to homogeneity from the culture filtrate of the mushroom *Termitomyces clypeatus*. The enzyme had optimum activity at pH 5.0 and temperature 65°C and was stable up to 60°C and within pH 2-10. Among the substrates tested, p-nitrophenyl- β -D-glucopyranoside and cellobiose was hydrolysed best by the enzyme. K_m and V_m values for these substrates were 0.5, 1.25 mM and 95, 91 μ mol/min per mg, respectively. The enzyme had low activity towards gentiobiose, salicin and β -methyl-D-glucoside. Glucose and cellobiose inhibited the β -D-glucosidase (PNPGase) activity competitively with K_i of 1.7 and 1.9 mM, respectively. Molecular mass of the native enzyme was approximated to be 450 kDa by HPLC, whereas sodium dodecyl sulphate polyacrylamide gel electrophoresis indicated a molecular mass of 110 kDa. The high molecular weight enzyme protein was present both intracellularly and extracellularly from the early growth phase. The enzyme had a pI of 4.5 and appeared to be a glycoprotein.

L3 ANSWER 5 OF 5 BIOSIS COPYRIGHT 2001 BIOSIS

Full Text	Citing References
-----------	-------------------

AN 1990:196921 BIOSIS

DN BA89:103592

TI BETA GLUCOSIDASE PRODUCTION BY THE MYCELIAL CULTURE OF THE MUSHROOM
 TERMITOMYCES-CLYPEATUS.
 AU SENGUPTA S; SENGUPTA S
 CS INDIAN INST. CHEM. BIOL., 4 RAJA S.C. MULLICK RD., CALCUTTA-700032,
 SO ENZYME MICROB TECHNOL, (1990) 12 (4), 309-314.
 CODEN: EMTED2. ISSN: 0141-0229.
 FS BA; OLD
 LA English
 AB Extracellular cellobiase activity was detected in the mycelial cult
 of the mushroom T. clypeatus with different mono-, di-, and
 polysaccharides as carbon source. Higher carbohydrate (2-5%) in the
 strongly repressed enzyme production without inhibiting growth rate
 the other hand, nonglucose monosaccharides also could not improve
 extracellular enzyme activity. Casein hydrolysate (CH) in the medi
 (w/v) concentration largely improved enzyme titer irrespective of c
 source (glucose, xylose, cellobiose, starch) used. Extracellular ac
 also appeared in high carbohydrate media in the presence of casein
 hydrolysate. The kinetics of extra- and intracellular production of
 enzyme in cellobiose (CB) medium, with or without CH, indicated
 extracellular and growth-dependent production of the enzyme. A maxi
 intracellular level of 8% of the total cellobiase was measured at 1
 late phase of growth in CB medium. CH had no effect on pH, temperat
 optima, and thermal stability of the enzyme produced in different
 carbohydrate-containing media. T. clypeatus did not liberate any
 proteinase in the presence or the absence of CH. Thus CH appeared r
 improve enzyme titer by repressing any proteinase or stabilizing er
 activity liberated in CH-free medium. It was therefore suggested tha
 constitutive production of cellobiase by T. clypeatus was under
 catabolic repression and CH probably released the repression to som
 extent. The β -glucosidase activity of the culture filtrate on
 p-nitrophenyl- β -D-glucose (pNPG), β -methyl-D-glucoside, and
 cellobiose had identical pH and temperature optima at 5° C and
 65° C, respectively. The enzyme had higher affinity for
 aryl- β -D-glucose, while β -CH₃-D-glucoside was a very poor
 substrate for the enzyme. The activity of the enzyme was readily ir
 by glucose, whereas glucose analogues or any other related sugars c
 have any appreciable inhibitory activity.

=>